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Asymmetric synthesis of (S)-(+)-carnitine and analogs

Rajendra P. Jain and Robert M. Williams*

Department of Chemistry, Colorado State University, Fort Collins, CO 80523, USA Received 12 February 2001; revised 23 March 2001; accepted 13 April 2001

Abstract—A general asymmetric route to enantiomerically pure (S)-(+)-carnitine and analogs has been investigated that involves monoaddition of organometallic reagents to the lactone carbonyl group of (5R,6S)-4-(benzyloxycarbonyl)-5,6-diphenyl-2,3,5,6-tetrahydro-4H-1,4-oxazin-2-one and Lewis acid promoted stereoselective allylation of the resulting hemiacetals. The diastereomerically pure allyl oxazines thus obtained were readily converted into enantiomerically pure (S)-(+)-carnitine and two substituted analogs. © 2001 Elsevier Science Ltd. All rights reserved.

The asymmetric synthesis of carnitine and its analogs has attracted considerable interest in recent years due to their important biological activities. (R)-(-)-carnitine plays an important role in biochemical pathways for β -oxidation of fatty acids and is involved in other important metabolic functions, both as free carnitine and as acyl carnitines. In addition, it has been found that this substance has clinical applications as a hypolipidemic agent in hemodialysis patients as well as in the treatment of myocardial ischemia and seizure. Interestingly, the antipode, (S)-(+)-carnitine acts as a competitive inhibitor of carnitine acyltransferase causing depletion of carnitine.

Several approached have been reported in the literature for the synthesis of carnitine and its analogs including asymmetric syntheses, 7 utilization of chiral starting materials, 8 chemical resolution, 9 enzymatic or microbial techniques 10 and others. 11 Herein, we report a general method for the synthesis of (S)-(+)-carnitine and its analogs using commercially available (5R,6S)-4-(benzyloxycarbonyl)-5,6-diphenyl-2,3,5,6-tetrahydro-4H-1,4-oxazin-2-one (1) 12 as a starting material.

The approach that was examined involved the sequential nucleophilic addition of organometalic agents to the lactone carbonyl group of 1, with the objective of creating a quaternary stereogenic center with high diastereoselectivity. Initial investigations were conducted with MeLi as the first nucleophile. Thus, reaction of 1 with MeLi at -78° C in a variety of etheral solvents (Et₂O, THF, DME) resulted in the formation of the desired monoaddition product 2b as a minor product (<25%) with recovery of a significant amount of unreacted starting material (Scheme 1). This was rationalized due to the competing enolization of 1 as a result of abstraction of the α -hydrogen by the alkyl lithium. However, addition of anhydrous CeCl₃ proved beneficial and the reaction of 1 with MeLi/CeCl₃¹³ (THF, -78°C, 4 h) generated the desired hemiacetal **2b** in 73% yield as a mixture of diastereomers at C2. Reaction of 1 with n-BuLi under identical conditions generated 2c in 45% yield (74% yield of 2c on the basis of recovered starting material) obtained as a mixture of diastereomers at C2. Conversion of 1 into the unsubstituted acetoxy hemiacetal 2a was achieved by reaction with DIBAL-H (CH₂Cl₂, −78°C, 30 min) followed by acetylation of the intermediate

Scheme 1.

Keywords: amino acids; carnitine; asymmetric synthesis.

^{*} Corresponding author. Tel.: +1-970-491-6747; fax: +1-970-491-5610; e-mail: rmw@chem.colostate.edu

Scheme 2.

hemiacetal (Ac₂O, Et₃N, DMAP, CH₂Cl₂, 0°C-rt, 12 h), as reported previously by our group¹⁴ (Scheme 1).

Reaction of **2b** and **2c** with allyltrimethylsilane/TiCl₄ (CH₂Cl₂, -78° C, 1 h) proceeded with a high degree of diastereoselectivity to generate the desired coupling products **3b** (67% yield) and **3c** (65% yield) as single diastereomers (as evidenced by 1 H NMR analysis of the crude reaction products). The relative stereochemistry of the newly created stereogenic center in **3b** and **3c** was determined as 'S' by 1 H NMR NOE measurements on **3b** and **3c**. Conversion of **2a** to **3a** was achieved by treatment with allyltrimethylsilane/BF₃·Et₂O (CH₃CN, -15 to -20° C, 30 min) as reported previously 14 (Scheme 2).

Modeling of the conformation of the putative oxocarbenium ion intermediate that is presumed to result from Lewis-acid-mediated removal of the acetoxy group from **2b** and **2c**, reveals that the oxazine ring adopts a boat-like conformation placing the phenyl ring adjacent to the ring nitrogen atom in *pseudo*-equatorial disposition mandating that the phenyl ring adjacent to the ring oxygen atom adopt an axial orientation (Fig. 1). The significantly less hindered

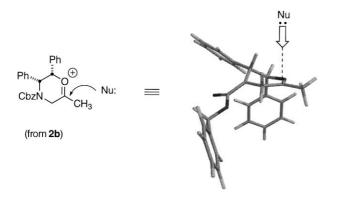


Figure 1. PM3 geometry optimization of the putative oxocarbenium ion intermediate using Spartan, C2 trivalent, overall molecular charge +1.

face of this intermediate thus suffers nucleophilic attack by the allyl group to furnish the *anti*-isomers **3b** and **3c** (Fig. 1).

Oxidative cleavage of the allylic double bond in $\bf 3b$ and $\bf 3c$ with O₃ (MeOH/CH₂Cl₂, -78° C, then Me₂S) followed by treatment with PDC (DMF, rt, 24 h) cleanly generated the carboxylic acids $\bf 4b$ and $\bf 4c$ in excellent yields (88–91% over two steps). Conversion of $\bf 3a$ to $\bf 4a$ was achieved in a similar way as reported previously. Removal of the chiral auxiliary in $\bf 4a-c$ by hydrogenolysis (PdCl₂, 120 psi H₂, THF/H₂O, 75–80°C, 3 h) followed by treatment with DOWEX® 50WX2-100 ion-exchange resin generated the desired β -hydroxy- γ -amino acids $\bf 5a-c^{11a,15}$ in essentially quantitative yields (Scheme 3).

Reductive methylation of $\bf 5a-c$ with aqueous formaldehyde (10% Pd/C, 80 psi H₂, rt, 36 h) generated the desired dimethylamino compounds $\bf 6a-c$ in essentially quantitative yields. Quaternization of $\bf 6a-c$ with MeI (DMF, rt, 12 h) followed by treatment with Amberlite® IRA-400 (HO-) ion-exchange resin generated (S)-(+)-carnitine $\bf 7a$ ([α]_D²⁵=+28.1 (c 1, H₂O); lit. 9 c [α]_D=-30.9 (c 1, H₂O) for the enantiomer) and the carnitine analogs $\bf 7b$ 11a ([α]_D²⁵=+18.3 (c 1, H₂O)) and $\bf 7c$ ([α]_D²⁵=+4.6 (c 1, H₂O)) in 68–85% yields (Scheme 4).

In summary, enantioselective syntheses of (S)-(+)-carnitine and two analogs have been achieved by the sequential mono-addition of organometallic reagents to the lactone carbonyl of (5R,6S)-4-(benzyloxycarbonyl)-5,6-diphenyl-2,3,5,6-tetrahydro-4H-1,4-oxazin-2-one, followed by Lewis acid-promoted stereoselective allylation of the resulting hemiacetals. Since (5S,6R)-4-(benzyloxycarbonyl)-5,6-diphenyl-2,3,5,6-tetrahydro-4H-1,4-oxazin-2-one is also commercially available, ¹² the methodology described here provides convenient access to (R)-(-)-carnitine and the corresponding analogs in the (R)-enantiomer series. Current efforts in these laboratories are devoted to expanding this

Scheme 4.

methodology to preparation of other amino acids, peptide isosteres and alkaloids of biomedical significance.

1. Experimental

All reactions requiring anhydrous conditions were performed under a positive pressure of argon using oven dried glassware (120°C) that was cooled under argon. THF was distilled from sodium benzophenone ketyl and dichloromethane was distilled from CaH₂. Column chromatography was performed on Merck silica gel Kieselgel 60 (230–400 mesh). H NMR and H NMR spectra were recorded on a Varian 300 or 400 MHz spectrometer. Mass spectra were obtained on Fisons VG Autospec. IR spectra were recorded on a Perkin-Elmer 1600 series FT-IR spectrometer. Optical rotations were obtained on a Rudolph Research automatic polarimeter Autopol III. Compounds 2a, 3a, and 4a were prepared according to the reported procedures. 14

1.1. General

1.1.1. General procedure for addition of alkyl lithium reagents to of (5R,6S)-4-(benzyloxycarbonyl)-5,6-diphenyl-2,3,5,6-tetrahydro-4H-1,4-oxazin-2-one. To anhydrous cerium chloride¹³ (1.2 equiv.) at 0°C was added anhydrous THF with vigorous stirring and the resulting suspension was warmed to and stirred at ambient temperature for 12 h after which the mixture was cooled to -78° C and the alkyl lithium was added over a period of 5 min. The resulting yellow suspension was stirred at the same temperature for 30 min after which a solution of 1 in anhydrous THF was added dropwise over a period of 15 min. The mixture was stirred at the same temperature for 4 h, quenched with saturated ag. NH₄Cl and warmed to ambient temperature. Dilute HCl was added and the mixture was extracted with ethyl acetate. The combined organic layer was dried (Na₂SO₄) and evaporated under reduce pressure to furnish the crude product which was purified by flash chromatography on silica gel using petroleum ether/ethyl acetate mixtures as eluents.

1.1.2. (5R,6S)-2-Hydroxy-2-methyl-5,6-diphenyl-morpholine-4-carboxylic acid benzyl ester (2b). Prepared from anhydrous CeCl₃ (1.65 g, 6.7 mmol) in 20 mL THF, MeLi (1.4 M in Et₂O, 4.78 mL, 6.7 mmol) and 1 (2.12 g, 5.5 mmol) in 100 mL THF. Purification of the crude product by flash chromatography on silica gel (8:2 petroleum ether: ethyl acetate) furnished 1.6 g (73%) of 2b as a white foam (mixture of diastereomers at C2). ¹H NMR (300 MHz, CDCl₃, 300K) δ 7.50–7.00 (m, 15H), 5.80–5.00 (m, 4H), 4.13 and 4.03 (d, 1H, J=13.5 Hz), 3.22 (d, 1H, J=13.5 Hz), 2.69 and 2.57 (s, 1H), 1.68 and 1.64 (s, 3H); IR (CHCl₃)

3409, 1678 cm⁻¹; **HRMS** (FAB+) Calcd for $C_{25}H_{26}NO_4$ (m/z) 404.1861, found (m/z) 404.1865.

1.1.3. (*5R*,*6S*)-2-Hydroxy-2-butyl-5,6-diphenyl-morpholine-4-carboxylic acid benzyl ester (2c). Prepared from anhydrous CeCl₃ (0.99 g, 4 mmol) in 15 mL THF, *n*-BuLi (2.3 M in hexane, 1.74 mL, 4 mmol) and **1** (1.28 g, 3.3 mmol) in 50 mL THF. Purification of the crude product by flash chromatography on silica gel (8:2 petroleum ether: ethyl acetate) furnished 0.663 g (45%) of **2c** as a white foam (mixture of diastereomers at C2) and 0.509 g of unreacted **1** as a white solid (74% yield of **2c** on the basis of recovered **1**). ¹**H NMR** (300 MHz, CDCl₃, 300K) δ 7.50–7.10 (m, 15H), 5.75–5.10 (m, 4H), 4.13 and 4.03 (d, 1H, J= 13.5 Hz), 3.23 (d, 1H, J=13.5 Hz), 2.81 and 2.65 (s, 1H), 2.00–0.80 (m, 6H), 1.04 (t, 3H, J=7.2 Hz); **IR** (CHCl₃) 3416, 1679, 1604, 1585 cm⁻¹; **HRMS** (FAB+) Calcd for C₂₈H₃₂NO₄ (m/z) 446.2331, found (m/z) 446.2331.

1.1.4. General procedure for allylation of hemiacetals 2b and 2c. To a solution of 2b and 2c in anhydrous CH_2Cl_2 was added allyltrimethylsilane at $-78^{\circ}C$ followed by $TiCl_4$ (1 M solution in CH_2Cl_2) over a period of 15 min and the solution was stirred at $-78^{\circ}C$ for 1 h, after which it was quenched with saturated aq. NH_4Cl and warmed to ambient temperature. Water was added to dissolve the precipitated solids, and the solution was extracted with CH_2Cl_2 . The combined organic phase was dried and concentrated to furnish the crude product which was purified by flash chromatography on silica gel using petroleum/ethyl acetate mixtures as eluents.

1.1.5. (*2S*,5*R*,6*S*)-2-Methyl-2-(2-propenyl)-5,6-diphenyl-morpholine-4-carboxylic acid benzyl ester (3b). Prepared from 2b (1 g, 2.48 mmol), allyltrimethylsilane (3.94 mL, 24.8 mmol) TiCl₄ (1 M solution in CH₂Cl₂, 9.92 mL, 9.92 mmol) in dichloromethane (10 mL). Purification of the crude product by flash chromatography on silica gel (9:1 petroleum ether:ethyl acetate) furnished 0.710 g (67%) of 3b as a clear colorless gum. $[\alpha]_D^{25}$ = -133.78 (*c* 1.9, CHCl₃); ¹H NMR (300 MHz, DMSO-d₆, 393K) δ 7.35–7.02 (m, 15H), 5.84–5.70 (m, 1H), 5.45 (d, 1H, *J*=3.6 Hz), 5.32 (d, 1H, *J*=3.6 Hz), 5.20 (d, 1H, *J*=12.6 Hz),), 5.12 (d, 1H, *J*=12.6 Hz), 5.14–5.03 (m, 2H), 3.79 (d, 1H, *J*=13.5 Hz), 2.37 (dd, 1H, *J*=7.5, 14.7 Hz), 1.33 (s, 3H); **IR** (CHCl₃) 1697 cm⁻¹; **HRMS** (FAB+) Calcd for C₂₈H₃₀NO₃ (*m/z*) 428.2225; found (*m/z*) 428.2208.

1.1.6. (2*S*,5*R*,6*S*)-2-Butyl-2-(2-propenyl)-5,6-diphenyl-morpholine-4-carboxylic acid benzyl ester (3c). Prepared from 2c (0.250 g, 0.56 mmol), allyltrimethylsilane (0.89 mL, 5.6 mmol), TiCl₄ (1 M solution in CH₂Cl₂, 2.24 mL, 2.24 mmol) in dichloromethane (3 mL). Purification of the

crude product by flash chromatography on silica gel (9:1 petroleum ether:ethyl acetate) furnished 0.173 g (65%) of **3c** as a clear colorless gum. $[\alpha]_D^{25} = -94.3$ (c 1, CHCl₃); $^1\mathbf{H}$ **NMR** (300 MHz, DMSO-d₆, 393K) δ 7.37–7.05 (m, 15H), 5.85–5.71 (m, 1H), 5.51 (d, 1H, J=3.6 Hz), 5.38 (d, 1H, J=3.6 Hz), 5.23 (d, 1H, J=12.3 Hz),), 5.16 (d, 1H, J=12.3 Hz), 5.14–5.05 (m, 2H), 3.77 (d, 1H, J=13.8 Hz), 3.06 (d, 1H, J=13.8 Hz), 2.70 (dd, 1H, J=6.6, 14.7 Hz), 2.47 (dd, 1H, J=7.2, 14.7 Hz), 1.73–1.65 (m, 2H), 1.54–1.34 (m, 4H), 0.95 (t, 3H, J=7.5 Hz); **IR** (CHCl₃) 1693, 1639, 1604, 1585 cm⁻¹; **HRMS** (FAB+) Calcd for $C_{31}H_{36}NO_3$ (m/z) 470.2695; found (m/z) 470.2628.

- 1.1.7. General procedure for the oxidation of allyl oxazines 3b and 3c. To a solution of 3a and 3b in anhydrous MeOH and anhydrous CH₂Cl₂ was bubbled O₃ at -78°C until the solution turned blue. At this point Ar gas was bubbled through the solution for 5 min to remove excess of O₃. Me₂S was added and the mixture was warmed to and stirred at ambient temperature overnight. The residue obtained after the evaporation of solvent was taken up in water and extracted with CH₂Cl₂. The combined extracts were dried (Na₂SO₄) and evaporated to provide the crude aldehyde which was treated with PDC (3.5 equiv.) in anhydrous DMF for 24 h. Water was added followed by brine and the mixture was extracted with ethyl acetate. The combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated to furnish the crude product which was purified by flash chromatography on silica gel using petroleum ether/ethyl acetate mixtures as eluents.
- 1.1.8. (2S,5R,6S)-2-Methyl-2-(2-carboxyethyl)-5,6-diphenyl-morpholine-4-carboxylic acid benzyl ester (4b). Prepared from 3b (0.5 g, 1.17 mmol) in dichloromethane (10 mL) and MeOH (20 mL) to obtain 0.486 g of crude aldehyde which was treated with PDC (1.5 g, 3.98 mmol) in anhydrous DMF (4 mL) to obtain crude 4b which on purification by flash chromatography on silica gel (1:1 petroleum ether:ethyl acetate) furnished 0.46 g of 4b as a white foam (88% yield over two steps). $[\alpha]_D^{25} = -118.3$ (c 1, CHCl₃); ¹**H NMR** (300 MHz, DMSO-d₆, 373K) δ 11.80 (br, 1H), 7.37–7.07 (m, 15H), 5.48 (d, 1H, *J*=3.6 Hz), 5.43 (d, 1H, J=3.6 Hz), 5.21 (d, 1H, J=12.9 Hz), 5.14 (d, 1H, J=12.9 Hz),J=12.9 Hz), 4.01 (d, 1H, J=14.1 Hz), 3.04 (d, 1H, J=14.1 Hz) 14.1 Hz), 2.85 (d, 1H, J=14.4 Hz), 2.72 (d, 1H, J=14.4 Hz), 1.54 (s, 3H); **IR** (CHCl₃) 3500–2700 (br), 1704, 1603, 1585 cm⁻¹; **HRMS** (FAB+) Calcd for C₂₇H₂₈NO₅ (m/z) 446.1967; found (m/z) 446.1969.
- **1.1.9.** (2S,5R,6S)-2-Butyl-2-(2-carboxyethyl)-5,6-diphenyl-morpholine-4-carboxylic acid benzyl ester (4c). Prepared from 3c (0.5 g, 1.06 mmol) in dichloromethane (10 mL) and MeOH (20 mL) to obtain 0.482 g of crude aldehyde which was treated with PDC (1.5 g, 3.98 mmol) in anhydrous DMF (4 mL) to obtain crude 4c which on purification by flash chromatography on silica gel (1:1 petroleum ether: ethyl acetate) furnished 0.47 g of 4c as a white foam (91% yield over two steps). $[\alpha]_D^{25} = -127.8$ (c 1, CHCl₃); ¹H NMR (300 MHz, DMSO-d₆, 373K) δ 11.80 (br, 1H), 7.37–7.07 (m, 15H), 5.51 (d, 1H, J=4.2 Hz), 5.45 (d, 1H, J=4.2 Hz), 5.21 (d, 1H, J=12.3 Hz),), 5.14 (d, 1H, J=12.3 Hz), 4.01 (d, 1H, J=13.5 Hz), 3.06 (d, 1H, J=13.5 Hz), 2.79 (s, 2H), 1.94–1.87 (m, 2H), 1.60–1.34

- (m, 4H), 0.95 (t, 3H, J=7.5); **IR** (CHCl₃) 3400–2800 (br), 1704, 1604, 1581 cm⁻¹; **HRMS** (FAB+) Calcd for $C_{30}H_{34}NO_5$ (m/z) 488.2436; found (m/z) 488.2433.
- **1.1.10.** General procedure for the hydrogenolysis of carboxylic acids 4a-c. A solution of 4a-c in THF: water (6:4, v/v) was hydrogenated with PdCl₂ (3 equiv.) at 75–80°C and 120 psi of H₂ for 3 h. The mixture was then cooled to ambient temperature, the catalyst was removed by filtration through a plug of Celite and the filtrate concentrated and triturated with Et₂O. The residue thus obtained was dissolved in water and passed through DOWEX[®] 50WX2-100 ion exchange resin and the resin was washed with distilled water. Elution of the resin with 2% aq. NH₄OH furnished the amino acids 5a-c which were pure by ¹H nmr analysis.
- **1.1.11.** (*S*)-4-Amino-3-hydroxybutanoic acid (5a). Prepared by hydrogenolysis of **4a** (0.12 g, 0.28 mmol) with PdCl₂ (0.15 g, 0.84 mmol) in THF (6 mL): water (4 mL) to provide 0.031 g (93%) of **5a** as a white amorphous solid after ion exchange chromatography. $[\alpha]_{\bf D}^{25}$ =+19.1 (*c* 1, H₂O); lit. $[\alpha]_{\bf D}^{25}$ =+20.7 (*c* 1.9, H
- **1.1.12.** (*S*)-4-Amino-3-methyl-3-hydroxybutanoic acid (**5b**).
 Prepared by hydrogenolysis of **4b** (0.13 g, 0.29 mmol) with PdCl₂ (0.16 g, 0.9 mmol) in THF (6 mL): water (4 mL) to provide 0.038 g (97%) of **5b** as a white amorphous solid after ion exchange chromatography. [α]_D²⁵=+8.4 (*c* 1, H₂O);
 H NMR (300 MHz, D₂O) δ 3.06 (d, 1H, *J*=12.9 Hz), 3.00 (d, 1H, *J*=12.9 Hz), 2.46 (s, 2H), 1.29 (s, 3H);
 H NMR (75 MHz, D₂O) δ 174.4, 64.4, 43.5, 41.8, 19.8; IR (KBr) 3700–2500 (br), 1638, 1561 cm⁻¹; HRMS (FAB+) Calcd for C₅H₁₂NO₃ (*m/z*) 134.0817; found (*m/z*) 134.0822.
- **1.1.13.** (*S*)-4-Amino-3-butyl-3-hydroxybutanoic acid (5c). Prepared by hydrogenolysis of 4c (0.13 g, 0.26 mmol) with $PdCl_2$ (0.15 g, 0.84 mmol) in THF (6 mL): water (4 mL) to provide 0.047 g (99%) of 5c as a white amorphous solid after ion exchange chromatography. $[\alpha]_D^{25} = +3.1$ (*c* 1.4, H₂O); ¹H NMR (300 MHz, D₂O) δ 3.13 (d, 1H, J = 9.9 Hz), 3.06 (d, 1H, J = 9.9 Hz), 2.48 (s, 1H), 1.60–1.56 (m, 2H), 1.34–1.29 (m, 4H), 0.88 (t, 3H, J = 5.4 Hz); ¹³C NMR (75 MHz, D₂O) δ 173.7, 65.4, 41.2, 38.2, 32.3, 19.5, 17.1, 7.8; IR (KBr) 3500–2400 (br), 1628, 1560 cm⁻¹; HRMS (FAB+) Calcd for $C_8H_{18}NO_3$ (m/z) 176.1286; found (m/z) 176.1285.
- **1.1.14.** General procedure for conversion of amino acids **5a-c** into **7a-c.** A solution of amino acids **5a-c** in distilled water was hydrogenated over 10% Pd/C in the presence of excess of 37% aq. formaldehyde at 80 psi of H₂ and ambient temperature for 36 h. The catalyst was removed by filtration through Celite and was washed with hot distilled water. The combined aqueous layer was passed through DOWEX[®] 50WX2-100 ion exchange resin and the resin was washed with distilled water. Elution of the resin with 2% aq. NH₄OH furnished the amino acids **6a-c** which were pure by ¹H NMR analysis. The amino acids **6a-c** were stirred with MeI in anhydrous DMF for 12 h at ambient temperature.

The solvent was removed under reduced pressure (2 mm) at ambient temperature to obtain the crude product which was dissolved in distilled water and passed through Amberlite[®] IRA-400 (OH) ion exchange resin eluting with water to furnish the amino acids 7a-c.

1.1.15. (S)-(+)-Carnitine (7a). Prepared by hydrogenation of **5a** (0.03 g, 0.25 mmol) in distilled water (4 mL) and formaldehyde (37 wt. % solution in water, 0.5 mL, excess) with 10% Pd/C (0.01 g, 33% w/w) to provide 0.037 g (99%) of **6a** as a white amorphous solid after DOWEX[®] 50WX2-100 ion exchange chromatography. ¹H NMR (300 MHz, D₂O) δ 4.40–4.26 (m, 1H), 3.22–3.10 (m, 2H), 2.90 (s, 6H), 2.39 (d, 2H, J=6.3 Hz). Treatment of **6a** (0.035 g, 0.23 mmol) with MeI (1 mL) and DMF (1 mL) followed by ion exchange chromatography using Amberlite® IRA-400 (OH) ion exchange resin furnished 0.03 g (78%) of **7a** as a clear colorless gum. $[\alpha]_D^{25}$ =+28.1 (c 1, H₂O); lit. $[\alpha]_D$ =-30.9 (c 1, H₂O) for the enantiomer; $[\alpha]_D$ + NMR (300 MHz, D₂O) $[\alpha]_D$ +0.0 for the enantiomer; $[\alpha]_D$ +1 NMR (300 MHz, D₂O) $[\alpha]_D$ +1.0 for the enantiomer; $[\alpha]_D$ +1.0 Hy, 2.22 (s, 9H), 2.50–2.36 (m, 2H).

1.1.16. (S)-3-Hydroxy-3-methyl-4-(trimethylammonio)butanoic acid (7b). 11a Prepared by hydrogenation of 5b (0.03 g, 0.22 mmol) in distilled water (4 mL) and formaldehyde (37 wt. % solution in water, 0.5 mL, excess) with 10% Pd/C (0.01 g, 33% w/w) to provide 0.036 g (99%) of **6b** as a white amorphous solid after DOWEX® 50WX2-100 ion exchange chromatography. ¹H NMR (300 MHz, D₂O) δ 3.32 (d, 1H, J=13.8 Hz), 3.23 (d, 1H, J=13.8 Hz), 2.95(s, 3H), 2.93 (s, 3H), 2.57 (s, 2H), 1.31 (s, 3H); ¹³C NMR (75 MHz, D₂O) δ 173.8, 64.8, 61.9, 43.0, 4 1.0, 21.8. Treatment of $\mathbf{6b}$ (0.030 g, 0.18 mmol) with MeI (1 mL) and DMF (1 mL) followed by ion exchange chromatography using Amberlite® IRA-400 (OH) ion exchange resin furnished 0.028 g (85%) of **7b** as a white amorphous solid. $[\alpha]_{D}^{25} = +18.3 (c 1, H_2O); {}^{1}H NMR (300 MHz, D_2O) \delta$ 3.52 (d, 1H, J=14.1 Hz), 3.47 ((d, 1H, J=14.1 Hz), 3.25(s, 9H), 2.45 (s, 2H), 1.46 (s, 3H); ¹³C NMR (75 MHz, D₂O) δ 173.2, 66.5, 65.3, 50.0, 42.6, 21.7; **IR** (KBr) 1655, 1596 cm^{-1} ; **HRMS** (FAB+) Calcd for $C_8H_{18}NO_3$ (*m/z*) 176.1286; found (*m/z*) 176.1284.

(S)-3-Hydroxy-3-butyl-4-(trimethylammonio)butanoic acid (7c). Prepared by hydrogenation of 5c (0.03 g, 0.25 mmol) in distilled water (4 mL) and formaldehvde (37 wt. % solution in water, 0.5 mL, excess) with 10% Pd/C (0.01 g, 33% w/w) to provide 0.034 g (97%) of **6c** as a white amorphous solid after DOWEX® 50WX2-100 ion exchange chromatography. ¹H NMR (300 MHz, D₂O) δ 3.29 (d, 1H, J=14.1 Hz), 3.17 (d, 1H, J=14.1 Hz), 2.90 (s, 6H), 2.57 (s, 2H), 1.60–1.48 (m, 2H), 1.36–1.20 (m, 4H), 0.85 (t, 3H, J=6.0 Hz); ¹³C NMR (75 MHz, D₂O) δ 173.9, 67.1, 61.2, 41.0, 40.4, 35.0, 20.1, 17.8, 8.7. Treatment of **6c** (0.030 g, 0.14 mmol) with MeI (1 mL) and DMF (1 mL) followed by ion exchange chromatography using Amberlite® IRA-400 (OH) ion exchange resin furnished 0.022 g (68%) of 7c as a clear colorless gum. $\left[\alpha\right]_{D}^{25}$ +4.6 (c 1, H₂O); ¹H NMR (300 MHz, D₂O) δ 3.52 (d, 1H, J=10.5 Hz), 3.46 (d, 1H, J=10.5 Hz), 3.26 (s, 9H),

2.52 (s, 2H), 1.80–1.64 (m, 2H), 1.36–1.26 (m, 4H), 0.87 (t, 3H, J=6.9 Hz); ¹³C NMR (75 MHz, D₂O) δ 173.4, 67.6, 65.0, 50.1, 38.8, 34.5, 19.8, 17.0, 7.8; IR (KBr) 1587 cm⁻¹; HRMS (FAB+) Calcd for C₁₁H₂₄NO₃ (m/z) 218.1756; found (m/z) 218.1753.

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References

- Ferrari, R.; Di Mauro, S.; Sherwood, G. L-Carnitine and its Role in Medicine: from Function to Therapy; Academic: San Diego, 1992.
- De Simone, C.; Famularo, G. Carnitine Today; Springer Verlag: Heidelberg, 1997.
- 3. Guarnieri, G. F.; Ranieri, F.; Toigo, G.; Vasile, A.; Ciman, M.; Rizzoli, V.; Moracchiello, M.; Campanacci, L.; Am *J. Clin. Nutr.* **1980**, *33*, 1489.
- 4. Woster, P. M.; Murray J. Med. Chem. 1986, 29, 865.
- Cavazza, C. Eur. Pat. Appl. EP 637449, 1995; Chem. Abstr. 1995, 122, 205212.
- (a) Vary, T. C.; Neely, J. R. Am. J. Physiol. 1982, 242, H585.
 (b) Bressler, R.; Brendel, K. J. Biol. Chem. 1966, 241, 4092.
- (a) Kitamura, M.; Ohkuma, T.; Takaya, H.; Noyori, R. Tetrahedron Lett. 1988, 29, 1555. (b) Takeda, H.; Hosokawa, S.; Aburatani, M.; Achiwa, K. Synlett 1991, 193. (c) Kolb, H. C.; Bennani, Y. L.; Sharpless, K. B. Tetrahedron: Asymmetry 1994, 4, 133.
- (a) Bellamy, F. D.; Bondoux, M.; Dodey, P. *Tetrahedron Lett.* 1990, 31, 7323. (b) Bols, M.; Lundt, I.; Pedersen, C. *Tetrahedron* 1992, 48, 319. (c) Kabat, M. M.; Daniewski, A. R.; Burger, W. *Tetrahedron: Asymmetry* 1997, 8, 2663.
- (a) Voeffray, R.; Perlberger, J.-C.; Tenud, L.; Gosteli, J. Helv. Chim. Acta 1987, 70, 2058. (b) Marzi, M.; Minetti, P.; Moretti, G.; Tinti, M. O.; De Angelis, F. J. Org. Chem. 2000, 65, 6766.
- (a) Kaneko, T.; Yoshida, R. Bull. Chem. Soc. Jpn. 1962, 35, 1153.
 (b) Zhou, B.; Gopalan, A. S.; VanMiddlesworth, F.; Shieh, W.-R.; Sih, C. J. J. Am. Chem. Soc. 1983, 105, 5925.
 (c) Gopalan, A. S.; Sih, C. J. Tetrahedron Lett. 1984, 25, 5235.
 (d) Fuganti, C.; Grasselli, P. Tetrahedron Lett. 1985, 26, 101.
 (e) Bianchi, D.; Cabri, W.; Cesti, P.; Francalanci, F.; Ricc, M. J. Org. Chem. 1988, 53, 104.
 (f) Kasai, N.; Sakaguchi, K. Tetrahedron Lett. 1992, 33, 1211.
- (a) Comber, R. N.; Hosmer, C. A.; Brouillette, W. J. J. Org. Chem. 1985, 50, 3627. (b) Brouillette, W. J.; Saeed, A.; Abuelyaman, A.; Hutchison, T. L.; Wolkowicz, P. E.; McMillin, J. B. J. Org. Chem. 1994, 59, 4297.
- 12. The requisite diphenyloxazine and its antipode are commercially available from Aldrich Chemical Co., Catalog #331856 (CAS Registry #105228464); the antipode: catalog #331872 (CAS Registry #100516549).
- 13. Takeda, N.; Imamoto, T. Organic Syntheses 1998, 76, 228.
- 14. Aoyagi, Y.; Williams, R. M. Tetrahedron 1998, 54, 10419.
- 15. Lu, Y.; Miet, C.; Kunesch, N.; Poisson, J. E. *Tetrahedron: Asymmetry* **1993**, *4*, 893.